

seed it also takes 3 -4 weeks with that type medium or you can put a clear plastic cup for high humidity which might speed up the germination process a week earlier. I don't fertilize the seedlings but I treat with Superthrive solution once every two weeks. After they reach one-year old I don't use this hormone anymore since by now the seedlings have good roots. When they are three years old, they are transferred to pure live spagnum moss or 50-50 peat/sand mix.

I feel the key to growing these plants is filtered sunlight, good low conductivity water and allowing them a dormancy period. I leave the pot of seedlings in water all year. I spray-mist the seedlings twice a day: in the morning and in the evening. If they get any insect pests on them, I spray with Orthene pesticide which is very good for CP and doesn't harm the most sensitive plants.

Followup on Tuberos Drosera Propagation

by

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In CPN Vol 20, No 3, P. 68, I reported on my attempts at the propagation of tuberos droseras by detaching the newly emerging plant from the tuber at the start of the season.

It is now the end of the season, and my tuberos droseras have all been unpotted and stored in small plastic bags for the summer. Normally, I would leave the tubers in their pots over the dormant period, but as we will be moving before the next growing season, I have packed them ready to be moved.

On inspecting the pots, I found that I had varying results with my experiments. Some detached plants produced tubers, whereas others of the same species did not. I would suspect that it is necessary to detach the plants a little closer to the tuber, in order to remove more of the stolon connecting the plant to the tuber, but still leaving sufficient stolon with dormant buds on the tuber to allow a new plant to be produced by the tuber after disconnection of the previous one.

Another method that can also be successful is to detach the newly formed tuber from the plant just before the plant starts to die down. This method is probably most successful with noncolonising species, i.e. ones that form a single replacement tuber at the end of the season. Many colonising species die down before the new tuber is formed, relying on the succulent I underground rhizome to produce the tuber. The new tuber is removed from the plant as soon as it is of sufficient size, and treated as a dormant tuber. The plant is again repotted and kept growing with weekly foliar fertilizing, and a new but probably substantially smaller tuber should be produced.

As a result of my weekly fertilizing program, I found that I had a higher survival rate of tubers, a higher average multiplication rate, and a larger average size of tubers.

I would expect to have a higher percentage of plants flowering in the following growing season as a result of the increase in tuber sizes. I used an indoor plant food of the low nitrogen type. I mixed it at full strength as recommended for the watering of indoor plants, but applied the fertiliser as a foliar spray. The type of fertilizer (whether organic or chemical) does not seem critical, but a correct balance of major nutrients is important, and trace nutrients would be considered beneficial also. Be sure to use a low nitrogen type, not a high nitrogen type, which may not give such good results.

Something else that I discovered accidentally, was the ability of some species to produce additional tubers from a stolon. While I was aware that this often happens with growing plants, I was a little surprised at my findings when inspecting a tuber of *D. macrophylla*. I had packed the tuber with attached stolon before the stolon had died back, and when I next looked several weeks later, I found several small “seedling” tubers along the length of the stolon. Hopefully these will survive to produce new small plantlets in the new growing season.

CLODS, Collectors, and Pseudo-Environmentalists

by

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I hate broken crayons. I had my first set when I was a youngster. I cherished them. When not using them in my colouring books or on the dining room walls, I lovingly kept them stored in their carton, arranged by colour. I stewarded my later and larger sets even more carefully. Every crayon was in excellent shape; I was disgusted if any were broken, dull, or embarrassingly short. Complete sets were nearly mandatory. I had the mentality of a collector even as a child.

I am still a collector, but now of books, music, and gargoyles. But my collector's personality makes its most breathtaking appearance in my relationship with carnivorous plants. When my carnivorous plant menagerie was still in its adolescence, I was not a collector so much as an accumulator, a packrat. I wanted every plant I did not have and would never have dreamt of parting with a species (although I constantly exchanged propagules with my trading comrades). With time I became more discriminating, and now I only grow *Utricularia*, *Genlisea*, *Sarracenia*, and several other favorite plants. (All right, I admit that about 150 species are more than several, but I do grow fewer extras than I once did. Really!)

A ritual performed by most plant traders, and especially those haunted by the hobgoblin that turns them into collectors, is the design and creation of an inventory of the plants in the trader's possession. Yes, I have built such a list, and to see it is to glimpse the raw mania of a collector's mentality. For each plant I grow, my list tabulates its size, trading availability, a short description of the plant's characteristics, and any other minutiae I deem important. All these data are compulsively arranged in a strict format—an inconsistency in any parenthesis or comma used to delimit data fields would be intolerable. Spelling errors would be sacrilegious.

My *Sarracenia* collection, a modern day set of crayons, is complete, or so it was until a few years ago. Consider the genus—it is easy to have a complete collection of these plants. The usual laundry list of species and subspecies includes only a dozen or